DIAGNOSIS AND PREVENTION OF INFECTION BY PHLEBOTOMUS FEVER GROUP VIRUSES

ANNUAL REPORT

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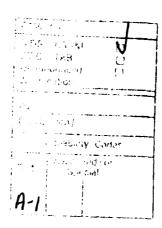
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I. SUMMARY

The objectives of the contract are to develop new diagnostic procedures and new vaccine strategies pertinent to selected Phlebotomus fever (PHL) group viruses, viruses that constitute the *Phlebovirus* genus of the negative-sense RNA virus family Bunyaviridae [Bishop et al., 1980]. This genus of viruses includes members that are known animal [including human] pathogens and that are therefore of consequence to military and civilian personnel in particular regions of the world [Bishop & Shope, 1979].

The development of new diagnostic tools for the specific identification of PHL group viruses is being investigated using eukaryotic expression vectors and cDNA clones of particular phlebovirus genes that are available to us. Two expression systems are under study, vaccinia and the insect specific baculovirus expression vectors. So far, it has proven difficult to express the complete M RNA gene products of Punta Toro (PT) phlebovirus using either vector. Recombinant vaccinia viruses have, however, been made that express part of the PT M RNA, producing the PT glycoproteins G1 and G2. The expressed products have been characterized. These studies were undertaken in conjunction with staff of Professor R.W.Compans of the University of Alabama in Birmingham. The expression of PT S RNA gene products supported by this contract and its immediate predecessor [DAMD-17-G-4035, see Overton et al., 1987] using Autographa californica nuclear polyhedrosis virus [AcNPV] baculovirus vectors is also reported.

The objective of developing new vaccine strategies for PHL group viruses is being realized through the characterization of an attenuated, candidate vaccine derivative of Rift valley fever [RVF] phlebovirus [the M12 mutant of RVFV isolate ZZ548]. This candidate vaccine was previously developed by staff of the Principal Investigator working at USAMRIID [Caplen et al., 1984]. In this reporting period the complete sequence of the viral M RNA of the M12 RVFV derivative has been obtained.

II. REPORT

A. Introduction

Our analyses have been directed towards developing new strategies for phlebovirus vaccine development. Initial studies, supported by prior U.S. Army Medical Research and Development contracts, were aimed at characterising the genome and structural components of phleboviruses. With the demonstration of a tripartite RNA genome for phleboviruses, genetic analyses were then undertaken to delineate the coding strategies of the viral RNA species and to determine if recombinant viruses could be obtained and used for vaccine purposes. By analyses of intertypic reassortant PT viruses we initially showed that the viral 7×10^5 dalton small [S] RNA species codes for the viral 26.9×10^3 dalton nucleocapsid [N] protein. The results of cloning and sequencing the S RNA of PT phlebovirus [Ihara et al., 1984] confirmed these data and showed that an open reading frame in the viral-complementary S RNA sequence coded for N [S mRNA]. A second open reading frame in the viral-sense strand coding for a 29.1×10^3 dalton non-structural protein, NS_S [Ihara et al., 1984] was also identified. The two subgenomic PT S induced mRNA species [one viral-complementary, the other viral-sense] were characterized with regard to their 3' and 5' end sequences and rates of synthesis in infected cells [Ihara et al., 1985a; Emery & Bishop, 1987].

The results of cloning and sequencing the $2x10^6$ dalton middle [M] size RNA of PT virus [Ihara $et\ al.$, 1985b] demonstrated that the PT M RNA codes for the $50\cdot70x10^3$ dalton viral glycoproteins, G1 and G2. A $30x10^3$ dalton non-structural protein, NS_M, that constitutes the amino terminal end of the PT glycoprotein precursor was also identified. The data showed that the order of the PT M gene products was NS_M-G1-G2. The PT M viral-complementary mRNA species were characterized [Ihara $et\ al.$, 1985a]. The results obtained for the PT M RNA correlated with the data reported by Collett and associates [1985] for RVFV M RNA and its gene products [RVFV M gene order: NS_M-G2-G1]. Although not proven, it is assumed that the phlebovirus $3x10^6$ dalton large [L] RNA species codes for the $200x10^3$ dalton large protein [a putative transcriptase component] found in viruses.

The genetic studies, including interference assays, reported from the work conducted in the previous contract, indicated that although intertypic reassortant viruses could be obtained, heterotypic virus interactions were not demonstrable (i.e., heterotypic phleboviruses did not interfere with each other and did not reassort their genomes in dual virus infections). Although not all phleboviruses were tested for genetic interactions, the results did not hold out much hope for this approach for vaccine development. Consequently, an alternative approach, that of developing candidate, including subunit, vaccines has been proposed for this contract. These objectives, together with research directed towards the

development of new and specific diagnostic reagents form the principal objectives of this contract.

B. Results from this Reporting Period.

The sequence of the MRNA of the M12 candidate vaccine of RVFV

Based on analyses of cDNA clones, the sequence of the MRNA of the ZZ501 strain of RVFV has been reported by Collett and associates (1985). In order to characterize the candidate RVFV vaccine that was produced from the related ZZ548 RVFV [Caplen et al., 1984], we have sequenced the M RNA of the M12 derivative. The sequence was obtained by analyses of λ cDNA clones. The data are shown in Fig. 1 [manuscript in preparation]. Based on the cDNA sequence analyses the RVFV M12 M RNA is deduced to be 3885 nucleotides long (mol. wt. 1.38x106, base composition: 27.3% A, 27.2% U, 25.4% G, 20.1% C) and has 3'- and 5'-terminal sequences that are complementary for some 9 residues. The viral RNA codes in its viral-complementary sequence for a single long gene product [the viral glycoprotein precursor] that is comprised of 1197 amino acids (130,000 daltons). The gene product is abundant in cysteine residues but has few potential asparagine-linked glycosylation sites. It is initiated by a methionine codon at nucleotide residues 21-23. These data are similar to those reported for RVFV ZZ501 M RNA [as ammended by recent studies at USAMRIID, J. Dalrymple, personal communication]. However, unlike the ZZ501 sequence, there is a second AUG codon [residues 9-11] in the viral-complementary sequence upstream of the M12 M gene product [Fig. 1]. It is followed by valine and histidine codons then a translation terminator [TAA, residues 18-20]. Even discounting this region the 5' non-coding region of the RVFV M viral-complementary RNA is short (20 nucleotides); the 3'-noncoding sequence is much longer (271 nucleotides). No other large open reading frame has been identified in either the viral, or viralcomplementary RVFV M RNA sequences.

Limited amino-terminal sequence analyses of the ZZ501 RVFV glycoproteins indicated the gene order and potential cleavage sites in the glycoprotein precursor. The data suggested the existence of a 23×10^3 dalton polypeptide (designated NS_M) in the glycoprotein precursor that precedes the RVFV G2 and G1 proteins (i.e., gene product order: NS_M-G2-G1). Examination of the sequence of the M12 gene product indicates that the ZZ548 M12 and ZZ501 sequences are closely related and that the amino terminal sequences of the viral

Fig. 1. Sequence of the M cDNA of RVFV M12

H V H * M Y V L T I L I S V L V C E A I I R V S L S S I R E E I C F G D S ACACAAAGATGATGATATATATATTITITATAACAATICTAACCTIGGTUTGTGAAGCGATTATTAGAGTGTCTCAAGCCCCACAAGAAGAAGACCTGCTTTGSTGAC1 10 20 30 40 50 60 70 80 90 100 110 120
T N P E H I E G A W D S L R E E E H P E E L S C S I S G I R E V K T S S D E L Y CCACTAACCCAGAGAGTGATGAAGGAGGTTAACACGAGGAGTTAT 130 140 150 160 170 180 190 200 210 220 230 240
RALKAIIAADGLNNITCHGKDPEDKISLIKGPPHKKRVGI ACAGGGCATTAAAAGCCATCATTGCTGCTGATGGCTTGAACAACATCACCTGCCATGGTAAGGGACCAAGATTTCCCTUATAAAGGGTCCTCTCAAAAAAGGGGGGTGGGGA 250 260 270 280 290 300 310 320 330 340 350 360
V R C E R R R D A K D I G R K T M A G I A M T V L P A L A V F A L A P V V F A E TAGTICGGTGTGAGGAGAGAGAGAGAGAGAGAGAGAGAGAGAGAG
DPHLRNRPGKGHNY'IDGHTQEDATCKPVTYAGACSSFDVL AAGACCCCCATCTCAGAAACAAGACCAGGAACAACTACATTGACGGGATGACCCAGATGCAAACCTGTGACATATGCTGGGGATGTAGCAGTTTTGATGTCT 490 500 510 520 530 540 550 560 570 580 590 600
LEKGKFPLFDSYAHHRTLLEAVHDT11AKADPP5CDLLSA TGCTTGAAAAGGGAAAATTTCCCCTTTTCCAGTCGTATGCTCATAGAACTCTACTAGAGGCAGTCACGACAAGGCTGATCCCACCTAGCTGTGACCTCTGAGTG 610 620 630 640 650 660 670 680 690 700 710 720
H G N P C H K E K L V H K T H C P N D Y G S A H H L N N D G K H A S V K C P P K CTCATGGGAACCCCTGCTGCTGAAAGAAATGGCTTCAGTCAG
Y G L T E D C N F C R D M Y G A S L K K G S Y P L D D L F C C S S E D D G S K L AGTATGSGCTCACTGAAGACTGCAACTTTTGTAGGCAGATGACAGGTGCTAGCCTGAAGAGGGGTCTTATCCTCCCAAGACTTGTTTTGTCAGTCA
K T K M K G V C E V G V O A L K K C D G O L S T A H E V V P F A V F K N S K K V JAAAAA-CAAAAAAGGGGTCTGCGAAGTGGGGGTTCAAGCACTCAAAAAGTGTGATGGCCAACTCAGCACTGCACGTGGGCCTTTGCAGTGTTTAAGAACTCAAAGAAGG 970 980 990 1000 1010 1020 1030 1040 1050 1060 1070 1060
V L D K L D L K 1 E E N L L P D S F V C F E H K G D Y K G T H D S G D 1 K R E L THATCTTGA:AAGCTTGACCTTAAGACTGAGGAGAATCTGCTACCAGACTCATTTGTCTGTTTCGAGGATAAGGGACAGTACAAAGGAACAATGGACTCTGGTCAGACTAAGAGGGAGC 1090 1100 1110 1120 1130 1140 1150 1160 1170 1180 1190 1200
K S F D I S D C P K I G G H G S K K C T G D A A F C S A Y E C T A D Y A N A Y C TCADARACCTITICATATCCTCAGTGCCCAAGATTGGAGGACATGGTAGAAAGCTATT 1210 1220 1230 1240 1250 1260 1270 1280 1290 1300 1310 1320
S H A N G S G I V D I U V S G V W K K P L C V G Y E R V V V K R E L S A K P I U GITCACATGCT=ATGGGTCAGGGATTGTGCAGATACAAGTATCAGGUSTCTGGAAGAAGCCTTTATGTGTAGGGTC NAAGAGTGGTTGTBAAGAGAGACTCTGTGCCAAGCCCATCC 1330 1340 1350 1360 1370 1380 1390 1400 1410 1420 1420 1420
R V E P C T T C I T K C E P H G L V V R S T G F K I S S A V A C A S G V C V T G AGAGAGTIGASCCTIGCACAACTIGTATAACCAAATGTGAGCCTCATGGATTGGTIGTCCGATCAACAGGGTTCAAGATATCATCAGCAGTTGCTTAGCGGAGTTTGCGTCACAG 1450 1460 1470 1480 1490 1500 1510 1520 1530 1540 1550 1560
S Q S P S T E I T L K Y P G I S Q S S G G D I G V H H A H D D Q S V S S K I V A GATCGCAGGGTCCTTCCACCGAGGTTACCACTCAGGGATCCCCAGGGATATCCCAGGGTTCTCTGGGGGGACACTGAGGGGTTCACGTCAGTTAGCTCCAGTTAGTCAGTT

Fig. 1 (Contd)

Y S T Y L M L L I V S Y A S A C S E L I D A S S R I T T C S T E G V N T K C R GITATAGCACCTGATGTTATATGATGCTCCATATGCATCAGCATGTTCAGAACTGATTCAGGCAAGCTCCAGAATCACCACTTGCTCTACAGAGGGTGTTAACACCAAGTGTA 2050 2060 2070 2080 2090 2100 2110 2120 2130 2140 2150 2160 L S G T A L I R A G S V G A E A C L M L K G V K E D G T K F L K L K T V S S I GACTGTCTGGCACAGCATTGATCAGAGCAGGGTCGATTGGGGCAGAGGCTTGTTTGATGTTGAAGTGAAGCAAGTTCTTAAAGTTAAAACTGTCTAAAGTG 2170 2180 2190 2200 2210 2220 2230 2240 2250 2260 2270 2270 L G A C S S R F T N H G S V S L S L D A E G I S G S N S F S F I E S P G K G Y A
ACTIGGGAGCATCATCTAGCCGTTTCACAAACTGGGGTCAGTTAGCCCTCACTGGACGCAAGGGATTICAGGGCCCAAATAGCTTTTCTTTCATTGAGGCCCAGGCAAAGGGTATG
2650 2660 2670 2680 2690 2700 2710 2720 2730 2740 2750 2760 I V D E P F S E I P R Q G F L G E I R C N S E S S V L S A H E S C L R A P N L I CAATTGITGATGAGCCATTCTCAGAAATTCCTCGGCAAGGGTTCTTGGGGGAGATCAGGGTGCAATTCAGGAUTCCTCAGTCCTGAGTGCTCATGAATCATCCCTTAGGGCACCAAACCTTA 2770 2780 2790 2800 2810 2820 2830 2840 2850 2860 2870 2870 2880 I A I D P F D D R R E A G G E S T V V N P K S G S W N F F D W F S G L H S W F G
GGATGGCCATTGGATGCATTGGATGATGGGGGGATCAGAGGTTGGGATCCAGAATCTGGATCTTGGATTTTGGATTTTGTTGATGTTTTTGGATCTCATGAGTTGGTTTG
3370 3380 3390 3400 3410 3420 3430 3440 3450 3400 3450

GCTGGGAATTAAACTAACTCTTTGAAGTTGCACCGGTCTTTGTGT 3850 3860 3870 3880

glycoproteins are conserved between them. As shown for PT and the RVFV ZZ501 glycoproteins there are multiple hydrophobic sequences in the M12 M gene product, including a 22-28 amino acid carboxy-proximal, hydrophobic region (G1). This hydrophobic sequence is followed by a 11-amino acid-terminal sequence that contains 3 charged (basic) amino acids. The size and constitution of the carboxy-terminal region is consistent with a transmembranal and anchor function for this glycoprotein in the viral envelope. Other regions of the glycoprotein precursor contain sequences of amino acids with a predominantly hydrophobic character. Their functions are unknown.

Comparison has been made between the reported sequence of ZZ501 M RNA (and its gene product as amended by recent data from USAMRIID) and that of the M12 mutant of ZZ548, recognising that ZZ501 is not the parent of the mutant. Between the two sequences there are XX nucleotide differences and YY amino acid changes. Considering the fact that the M12 virus was derived by consecutive high level mutagenesis of another RVFV strain, these differences are small. Comparison with the sequence of the parent ZZ548 is under analysis at the present time.

Expression of the PT N and NS proteins using baculovirus expression vectors

An essentially complete DNA copy of the ambisense S RNA species of PT phlebovirus [Ihara et al, 1984a] has been inserted in either orientation into Autographa californica nuclear polyhedrosis baculovirus (AcNPV) in lieu of the 5' coding region of the AcNPV polyhedrin gene [Overton et al., 1987]. The recombinant viruses were generated using what proved to be proficient, but non-optimal expression vectors [e.g., pAcRP6, see below]. The two types of recombinant viruses were used to infect Spodoptera frugiperda cells and the expressed PT viral proteins characterised (Overton et al., 1987). Recombinant AcNPV having the S DNA in one orientation expressed PT virus N protein in amounts estimated to represent some 50% of the infected cell extracts, whereas recombinants with the PT S DNA in the other orientation expressed the PT virus NSs protein in significantly lower quantities. Antisera that were monospecific with respect to each of the two PT proteins virus were raised in mice using the corresponding S. frugiperda infected cell extracts and were employed to identify N and NSs proteins in PT virus-infected Vero cells. These studies demonstrated a

low amount of PT NS_S protein in virions and virion-derived nucleocapsids (Overton et al., 1987).

The results reported previously by this laboratory have now been extended by insertion of each individual gene in a more efficient expression vector derived in our laboratory [pAcYMI, Matsuura et al., 1987]. The level of expression of the PT N protein was similar to that obtained with the pAcRP6 vector, however the level of expression of the NSs protein was considerably enhanced, approaching that of the N protein. When these analyses are complete they will be prepared for publication.

Expression of PT virus glycoproteins from partial cDNA clones inserted into a vaccinia vector

Partial cDNA clones of the PT M RNA were inserted into the genome of vaccinia virus so that the PT gene products were under the control of an early vaccinia promoter. The PT sequences included in the constructs represented the entire G1 and G2 coding sequences preceded by some 24-37 upstream amino acids [depending on the constructions]. Recombinant vaccinia viruses were prepared using vaccinia plasmid pSC11 and transfection of CV-1 cells previously infected with strain IHD-J of vaccinia virus. Recombinant viruses were selected by the appropriate procedures [Smith and Moss, 1983]. In CV-1 cells the recombinants expressed glycosylated derivatives of both PT G1 and G2 proteins. In Hela T4 + cells the glycoproteins were identified in the Golgi apparatus. These studies were undertaken in conjunction with Ms Y. Matsuoka and Professor R.W. Compans of the University of Alabama in Birmingham [Matsuoka, Ihara, Bishop & Compans, 1988].

C. Summary of Progress Report

The sequence of the M RNA of the M12 derivative of ZZ548 has been determined and compared to that of ZZ501. The data indicate that the RNA and its gene product are comparable for both viruses. Comparison with the sequence of the parent ZZ548 virus is in hand.

Expression of the PT N and NS_s proteins has been achieved using baculovirus expression vectors based on AcNPV. High level expression of both proteins has now been achieved. Expression of the PT M gene products, G1 and G2, in vaccinia vectors has also been achieved

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